

Transcriptional Control Within the Three-Dimensional Context of Nuclear Architecture: Requirements for Boundaries and Direction

Gary S. Stein,* André J. van Wijnen, Janet L. Stein, Jane B. Lian, Sandra McNeil, and Shirwin M. Pockwinse

Department of Cell Biology and Cancer Center, University of Massachusetts Medical School, Worcester, Massachusetts 01655

Abstract Evidence is accruing that the architectural organization of nucleic acids and regulatory proteins within the cell nucleus support functional interrelationships between nuclear structure and gene expression. The punctate distribution of several transcription factors has provided several paradigms for pursuing mechanisms that direct these regulatory proteins to subnuclear sites where gene activation or suppression occurs. Sequences that are necessary and sufficient to direct regulatory proteins to transcriptionally active nuclear domains have been identified. Mutations that disrupt intranuclear targeting signals lead to modified subnuclear distribution of transcription factors and aberrant expression in tumor cells. *J. Cell. Biochem. Suppl.* 32/33:24–31, 1999. © 1999 Wiley-Liss, Inc.

Key words: AML/CBFA; gene expression; intranuclear targeting; subnuclear domains

Transcriptional control requires linkage of signaling pathways that exchange regulatory information between the nucleus and cytoplasm as well as within the cell nucleus. Gene promoters provide regulatory infrastructure by functioning as blueprints for responsiveness to the flow of cellular regulatory signals. Selective utilization of the information supports transient developmental and long-term phenotypic expression of genes. As the regulatory sequences and factors that activate or suppress transcription are identified and characterized, several fundamental paradoxes surface. How can a gene with a single promoter meet the selective requirements for expression under diverse biological circumstances? How, with a limited repertoire of promoter elements and cognate regulatory factors, can a threshold be attained to initiate or downregulate transcription in nuclei of intact cells? How are the regu-

latory activities that reside at independent promoter domains integrated?

There is increasing acceptance that components of nuclear architecture support the organization and sorting of regulatory information in a manner that facilitates selective utilization. The primary level of nuclear organization, the representation and ordering of genes and promoter elements, provides alternatives for physiological control. The modular arrangement of regulatory elements and overlap of regulatory sequences within promoter domains and the multipartite composition of regulatory complexes increases options for responsiveness. Chromatin structure and nucleosome organization reduces distances between regulatory sequences, facilitates crosstalk between promoter elements, and renders elements competent for interactions with positive and negative regulatory factors. The components of higher-order nuclear architecture that includes nuclear pores, the nuclear matrix and subnuclear domains contribute to the subnuclear distribution and activities of genes and regulatory factors.

There is emerging recognition that nuclear structure and function are causally interrelated. With accruing evidence for organization

Grant sponsor: National Institutes of Health; Grant number: AR45688; Grant number: AR45689.

*Correspondence to: Authors at Department of Cell Biology and Cancer Center, University of Massachusetts Medical School, 55 Lake Avenue North, Worcester, MA 01655. E-mail: gary.stein@umassmed.edu

Received 27 September 1999; Accepted 30 September 1999

of nucleic acids and regulatory proteins into subnuclear domains that are associated with components of nuclear architecture, the perception of a dichotomy between nuclear architecture and control of gene expression is no longer justifiable. Rather, the challenges now being faced are to design experiments to define mechanisms that direct genes and regulatory factors to sites within the nucleus where localization integrates regulatory parameters of gene expression and establishes boundaries between regulatory complexes that are necessary for fidelity of regulatory activity.

In this article, we focus on the involvement of nuclear architecture in transcriptional control with the appreciation that there is compelling evidence for contributions of nuclear structure to the stringent regulation of DNA replication [Leonhardt et al., 1998; Ma et al., 1999; Wei et al., 1998; Lamond and Earnshaw, 1998], post-transcriptional processing of gene transcripts [Wei et al., 1999; Iborra et al., 1998; Carter et al., 1993; Clemson et al., 1996; Nickerson et al., 1995; Pombo and Cook, 1996; Chen et al., 1994; Jackson et al., 1998], as well as the import, retention and export of nucleic acids and regulatory proteins [Silver et al., 1984; Ullman et al., 1997].

LINKAGE OF NUCLEAR ARCHITECTURE WITH COMPONENTS OF GENE EXPRESSION: A CONCEPTUAL AND EXPERIMENTAL BASIS FOR BOUNDARIES AND DIRECTIONS

Evidence for Architectural Organization of Genes and Regulatory Factors

From an historical perspective, it was recognized at the beginning of the twentieth century that the genome is packaged as chromosomes, providing the first indication that regulatory information is organized as discrete structures within the cell nucleus. The observation that chromosomes are “decondensed” to “interphase chromatin” at the completion of mitosis strikingly indicated that remodeling of a prominent nuclear structure occurs during a biologically relevant transition. The reorganization of the nucleolus in a cell cycle-dependent manner similarly revealed the plasticity of ribosomal gene packaging. The unique organization of DNA in lampbrush and in polytene chromosomes exhibited specialized architectural features of gene organization to support specific requirements for gene expression under biological conditions. Chromosome fragmentation and reorganiza-

tion in cancer, and more recently in neurological disorders, provided evidence for interrelationships between altered chromosome organization when fidelity of gene expression is compromised.

The discovery 25 years ago that chromatin is organized into “beads-on-a-string” structures, designated nucleosomes that are structurally remodeled to accommodate requirements for transcription, further emphasized the extent to which architectural organization of genes is linked to functional activity. Characterization of higher-order chromatin organization also demonstrated the intricacies of gene packaging and chromatin remodeling that accommodate activation and suppression of expression [reviewed in Zlatanova and van Holde, 1992; Imbalzano, 1998; Ito et al., 1997; Giese et al., 1995]. Taken together, these findings establish interrelationships between structural properties of genes and chromosomes with competency for transcription. Equally important, the requirement for modifications that mediate regulation of chromosome and chromatin remodeling in response to a broad spectrum of conditions became apparent. The initial documentation that reversible post-translational modifications of histones that included acetylation and phosphorylation are associated with alterations in gene activity and with remodeling of chromosomes and chromatin provided a basis for exploring mechanisms that control chromosome, chromatin and nucleosome organization. The recent identification and characterization of proteins that catalyze histone acetylation, deacetylation, phosphorylation [reviewed in Davie and Chadee, 1998], as well as the SWI/SNF-related proteins [Peterson et al., 1998; Imbalzano, 1998; Kwon et al., 1994; Cote et al., 1994; Workman and Kingston, 1998] that facilitate chromatin remodeling, and potentially the accessibility of promoter sequences to regulatory factors, represent a new dimension in the control of the structure and activity of genes and promoter regulatory elements. Linkage of regulatory signaling pathways to enzyme activity that modulates gene, chromatin and chromosome organization can now be directly investigated. Additional levels of specificity are provided by structural modifications of gene promoters that influence competency for factor interactions. Simply stated, changes in the architectural properties of promoter elements determine effectiveness of gene regulatory se-

quences as substrates for interactions with regulatory factors. The regulatory and regulated components of chromatin remodeling and the rate-limiting steps in the relevant signaling cascades are being actively pursued and unquestionably will provide insight into gene regulatory mechanisms from structural and functional perspectives.

The Subnuclear Organization of Regulatory Proteins

There is longstanding and compelling evidence for the asymmetric organization of protein complexes within the nucleus. Coiled bodies [Smith et al., 1999], PML domains [Dyck et al., 1994; Grande et al., 1996], and SC35 domains [Carter et al., 1993; Clemson et al., 1996; Blencowe et al., 1994; Nickerson et al., 1995; Pombo and Cook, 1996] are examples of protein localization at specific intranuclear sites that have been associated with gene expression. Transactivation factors that include AML [Merriam et al., 1995; Zeng et al., 1997, 1998; Chen et al., 1998], ALL [Rogaia et al., 1997; Yano et al., 1997], steroid hormone receptors [Tang et al., 1998a; van Steensel et al., 1995; Htun et al., 1996; Stenoien et al., 1998], and RNA polymerase II [Kimura et al., 1999; Zeng et al., 1998; Wei et al., 1998] exhibit a punctate subnuclear distribution.

Colocalization of regulatory factors with RNA polymerase II and RNA transcripts strengthens linkages between the intranuclear organization of regulatory complexes and fidelity of gene expression [Ma et al., 1999; Wei et al., 1999; Zeng et al., 1998]. Qualitative and quantitative variations in the representation and subnuclear distribution of regulatory factors are correlative but substantive support for involvement of nuclear structure with transcriptional control. However, evidence for interrelationships between nuclear architecture and regulation of gene expression that is difficult to dismiss is provided by demonstrations that transformed and tumor cells exhibit striking modifications in the intranuclear distribution of transcription factors.

From a fundamental regulatory perspective, organization of sites of transcription as discrete nuclear domains [Wei et al., 1998; Lamond and Earnshaw, 1998; Jackson et al., 1998; Iborra et al., 1998] provides an architectural basis for intranuclear organization of regulatory factors that mediate transcriptional control. Associa-

tion of transcription factors with the nuclear matrix also invokes nuclear architecture in gene regulation providing a scaffold for the organization of transcriptional regulatory machinery and foci with concentrations of factors that can support initiation of transcription. Yet it is essential to understand mechanisms that direct regulatory factors to subnuclear sites as well as parameters of the assembly, activation and remodeling of regulatory domains.

MECHANISMS THAT TARGET REGULATORY FACTORS TO TRANSCRIPTIONALLY ACTIVE SUBNUCLEAR SITES

There has been extensive investigation of the exchange of regulatory information between the nucleus and cytoplasm. Both the morphology of nuclear pore complexes and mechanisms that mediate nuclear import and egress of nucleic acids, proteins and nucleoprotein complexes have been characterized. However, the intranuclear targeting of transcription factors is minimally understood. The recent observation that members of the AML transcription factors exhibit a punctate subnuclear distribution and are associated with the nuclear matrix provided the basis for experimentally addressing the targeting of regulatory proteins to architecturally-associated sites within the nucleus that support transcription [Zeng et al., 1997]. Biochemical and in situ immunofluorescence analysis of transcription factors established that a 31-amino acid segment, designated the nuclear matrix targeting signal (NMTS) near the C-terminus is necessary and sufficient to direct the regulatory proteins to a nuclear matrix-associated subnuclear site, where transcription occurs [Zeng et al., 1998; Zeng et al., 1997]. The NMTS functions autonomously and is independent of the DNA binding domain. Specificity of the intranuclear targeting signal in the AML transcription factors is indicated by a unique nucleotide sequence [Zeng et al., 1997] and a unique structure that was defined by x-ray crystallography [Tang et al., 1999; Tang et al., 1998b]. Specificity within the context of nuclear organization is supported by colocalization with activators and repressors of AML factor activity and the absence of colocalization with factors that have been shown to be functionally unrelated in biochemical assays. Targeting signals (Fig. 1) that have been characterized in the estrogen receptor and glucocorticoid receptor [Tang et al., 1998a; van Steensel et al., 1995;

Nuclear Matrix Targeting Signals in Gene Regulatory Factors				
Protein name	Transcription factor class	Gene regulatory function	NMTS domain	Structural domains
AML1/CBFA2	Runt homology domain	Hematopoiesis	31 amino acids	β strands connected by a U-shaped hinge
AML2/CBFA3	Runt homology domain	Intestinal development	27 amino acids	homologous to AML1 NMTS
AML3/CBFA1	Runt homology domain	Bone formation	37 amino acids	homologous to AML1 NMTS
YY1	Zn-finger domain	General	within 141 a.a.	region overlaps with Zn-finger DNA binding domain
PIT1	POU/homeodomain	Pituitary development	66 amino acids	helix-turn-helix DNA binding domain
GR	Ligand-dependent	Glucocorticoid signaling	bipartite	DNA binding domain/tau2 transactivation domain

Fig. 1. Nuclear matrix targeting signals in transcription factors. Nuclear matrix targeting signals (NMTS) have been identified in tissue-specific transcription factors (e.g., AML1/CBFA2, Pit1) [Zeng et al., 1997; Mancini et al., 1999], as well as other gene regulatory proteins (e.g., YY1, GR) [McNeil et al., 1998; Tang et al., 1998a]. Each of these proteins is representative of distinct classes of transcription factors with characteristic conserved protein domains (second column) and key roles in the biological

regulation of gene expression (third column). The approximate sizes of the NMTS (fourth column) and their structural properties (fifth column) are indicated. The crystal structure of the AML1 NMTS represents the first three-dimensional structure of a signal peptide uniquely dedicated to subnuclear targeting [Tang et al., 1998b, 1999]. The remaining NMTS sequences appear to be embedded at least in part within different types of DNA binding modules.

Htun et al., 1996] or in the PIT1 [Stenoien et al., 1998] and YY1 [Guo et al., 1995; McNeil et al., 1998] transcription factors do not share sequence homology with the AML transcription factors. Each exhibits independent subnuclear distributions reinforcing specificity for intranuclear trafficking. Taken together, a series of trafficking signals have been identified that are responsible for directing regulatory factors to nonoverlapping sites within the cell nucleus. Collectively, these transcription factors provide coordinates for studying the intranuclear organization of gene regulatory complexes and the necessary components of structure, as well as multidirectional crosstalk that is requisite for biological control.

REQUIREMENTS FOR INCREASED UNDERSTANDING OF NUCLEAR STRUCTURE-GENE EXPRESSION INTERRELATIONSHIPS

There is now a significant body of evidence that the architectural organization of nucleic acids and regulatory proteins within the cell nucleus supports functional interrelationships between nuclear structure and gene expression. The punctate distribution of several transcription factors has provided paradigms for pursuing mechanisms that direct these regulatory proteins to specific subnuclear sites that support the activation or suppression of gene expression.

It is becoming increasingly apparent that intranuclear trafficking of regulatory factors is

a multistep process. We are only at the initial stages of understanding the complexities that control each component of the process that is responsible for targeting transcription factors to subnuclear sites. However, biochemical and in situ analysis have shown that at least two trafficking signals appear to be required for subnuclear targeting of AML transcription factors; the first supports nuclear import (nuclear localization signal) and a second mediates association with the nuclear matrix (nuclear matrix targeting signal). The multiplicity of determinants for nuclear localization and alternative splicing of AML messenger RNA may provide the requisite complexity to support targeting to specific sites within the nucleus in response to diverse biological conditions. Furthermore, because gene regulation by AML transcription factors involves contributions by other regulatory proteins such as CBF β [Banerjee et al., 1996; Ogawa et al., 1993], ETS [Giese et al., 1995], and C/EBP [Zhang et al., 1996], AML may facilitate recruitment of these factors to nuclear matrix-associated sites.

There is growing appreciation for involvement of nuclear architecture in the dynamic and multidirectional exchange of gene transcripts and regulatory factors between the nucleus and cytoplasm, as well as between regions and structures within the nucleus. Targeting of regulatory factors to subnuclear sites that are dedicated to biological activity is not confined to transcription factors. It has been shown that DNA replication occurs at nuclear

matrix-associated foci [Berezney et al., 1996; Wei et al., 1998] and peptides have been identified that direct replication factors to sites within the nucleus where DNA synthesis occurs [Leonhardt et al., 1998].

It is difficult to separate nuclear structure and function arbitrarily or to distinguish the regulated and regulatory parameters of control. The challenges we now face are to further define the targeting of transcription factors and control that reside at the level of nuclear matrix-associated acceptor sites. Here it will be important to determine whether acceptor proteins are associated with a core filament structural

lattice or whether a compositely organized matrix of regulatory factors that contribute to nuclear morphology is dynamically assembled. The result will unquestionably be further insight into fundamental processes that are involved with directing components of gene expression to specific regions within the nucleus.

It is premature to define an inclusive pathway to account mechanistically for all steps in the targeting of proteins to subnuclear sites where they participate in regulatory activity. However, findings suggest that parameters of nuclear architecture functionally interface with components of transcriptional control (Fig. 2).

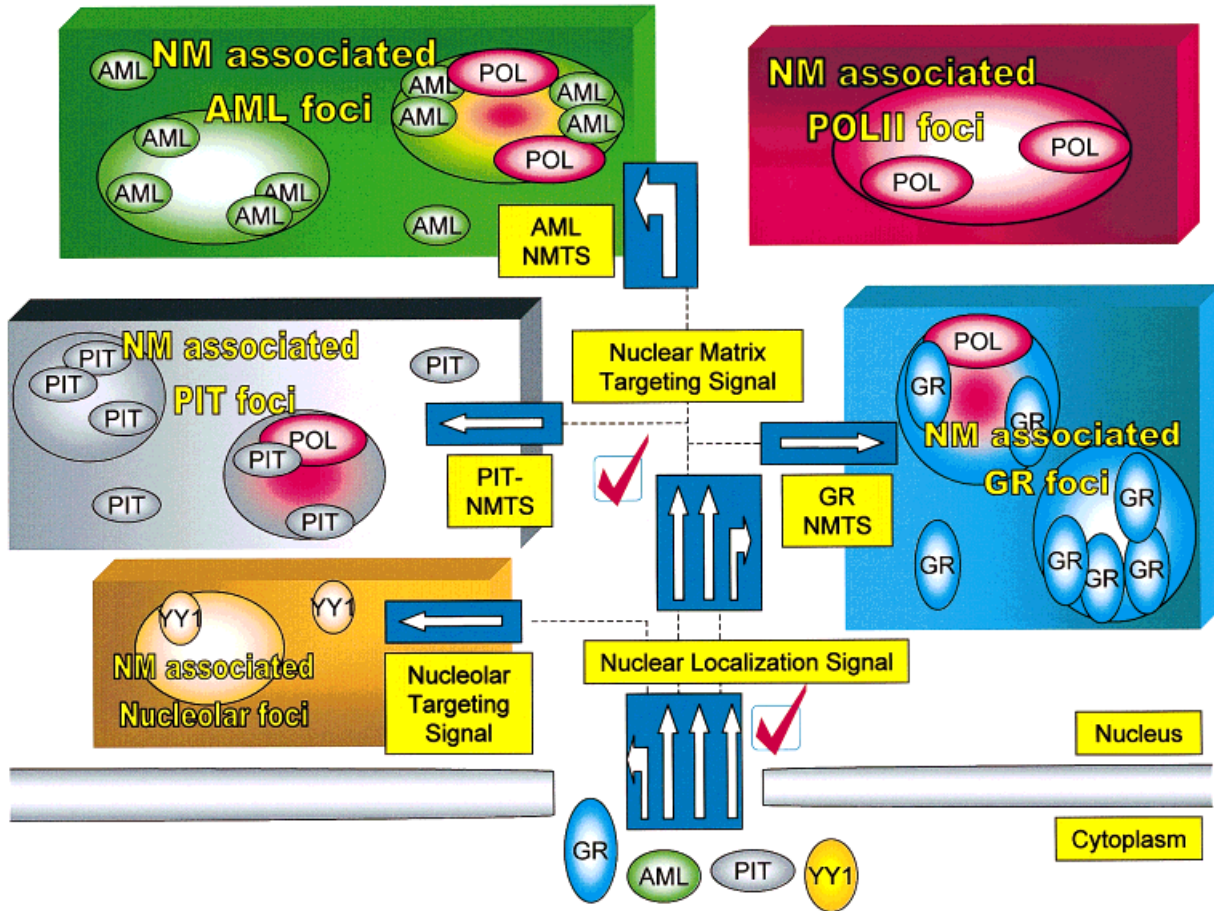


Fig. 2. Subnuclear targeting of transcription factors to gene regulatory sites. Transcription factors are imported into the nucleus using nuclear localization signals (NLS) that are structurally and functionally similar. Upon nuclear entry, regulatory factors are directed to subnuclear domains by intranuclear targeting signals (e.g., nuclear matrix targeting signal [NMTS]) (indicated in yellow boxes) that are structurally and functionally unique to specific transcription factors. Schematically illustrated is the trafficking (blue boxes with white arrows) of the transcription factors AML (small green ovals), GR (small blue

ovals), PIT (small gray ovals), YY1 (small orange ovals) to nuclear matrix-associated foci (large ovals), where genes are transcribed or repressed. Transcriptionally active domains containing RNA polymerase II are highlighted in red. Checkpoints designated in red indicate regulatory stages in nuclear import and subnuclear trafficking during which sorting and surveillance mechanisms are operative. The transcription factors indicated in the diagram represent paradigms for defining mechanisms that mediate sorting of regulatory proteins within the nucleus.

The involvement of nuclear matrix-associated transcription factors with recruitment of regulatory components to modulate transcription remains to be defined. The diversity of targeting signals must be established to evaluate the extent to which regulatory discrimination is mediated by encoded intranuclear trafficking signals. It will also be imperative to define, both biochemically and mechanistically, the checkpoints that are operative during subnuclear distribution of regulatory factors and the editing steps that are invoked to ensure the structural and functional fidelity of nuclear domains where replication and expression of genes occur. There is emerging recognition that placement of regulatory components of gene expression must be temporally and spatially coordinated to mediate biological control optimally. The consequences of breaches in nuclear structure-function interrelationships are observed in an expanding series of diseases that include cancer and neurological disorders. As the repertoire of architecturally associated factors and cofactors that modulate regulatory activity expand, there is increasing confidence in contributions of nuclear organization to transcriptional activity. Defining structure-function interrelationships is not an obstacle to controlling gene expression *in vivo*. Rather, mechanisms are emerging that use nuclear structure to organize nucleic acids and regulatory proteins in a manner that facilitates regulation of genes. It is difficult to dismiss the necessity for further characterization of mechanisms that direct regulatory proteins to specific sites within the nucleus and thereby be in the right place at the right time to participate in transcriptional control. Here, the boundaries between subnuclear domains are important to establish threshold concentrations of regulatory factors to support the isolation as well as integration of activities that accommodate specific biological requirements for expression of genes.

It is no longer justifiable to equate transcriptional control that can be reconstituted under cell free conditions with the activation and suppression of genes of nuclei of intact cells. Although components of regulatory mechanisms can be dissected *in vitro*, the biochemical and architectural complexities of the intact cell are necessary for the full complement of regulatory factors and integration of signaling mechanisms, to be operative for physiologically responsive control.

ACKNOWLEDGMENTS

Components of the work reported in this review were supported by National Institutes of Health grants AR45688 and AR45689. Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the National Institutes of Health. The authors are appreciative of the editorial assistance from Elizabeth Bronstein in the development of the manuscript.

REFERENCES

- Banerjee C, Hiebert SW, Stein JL, Lian JB, Stein GS. 1996. An AML-1 consensus sequence binds an osteoblast-specific complex and transcriptionally activates the osteocalcin gene. *Proc Natl Acad Sci USA* 93:4968-4973.
- Berezney R, Mortillaro M, Ma H, Meng C, Samarabandu J, Wei X, Somanathan S, Liou WS, Pan SJ, Cheng PC. 1996. Connecting nuclear architecture and genomic function. *J Cell Biochem* 62:223-226.
- Blencowe BJ, Nickerson JA, Issner R, Penman S, Sharp PA. 1994. Association of nuclear matrix antigens with exon-containing splicing complexes. *J Cell Biol* 127:593-607.
- Carter KC, Bowman D, Carrington W, Fogarty K, McNeil JA, Fay FS, Lawrence JB. 1993. A three-dimensional view of precursor messenger RNA metabolism within the mammalian nucleus. *Science* 259:1330-1335.
- Chen J, McKee MD, Nanci A, Sodek J. 1994. Bone sialoprotein mRNA expression and ultrastructural localization in fetal porcine calvarial bone: comparisons with osteopontin. *Histochem J* 26:67-78.
- Chen LF, Ito K, Murakami Y, Ito Y. 1998. The capacity of polyomavirus enhancer binding protein 2alphaB (AML1/Cbfa2) to stimulate polyomavirus DNA replication is related to its affinity for the nuclear matrix. *Mol Cell Biol* 18:4165-4176.
- Clemson CM, McNeil JA, Willard HF, Lawrence JB. 1996. XIST RNA paints the inactive X chromosome at interphase: evidence for a novel RNA involved in nuclear/chromosome structure. *J Cell Biol* 132:259-275.
- Cote J, Quinn J, Workman JL, Peterson CL. 1994. Stimulation of GAL4 derivative binding to nucleosomal DNA by the yeast SWI/SNF complex. *Science* 265:53-60.
- Davie JR, Chadee DN. 1998. Regulation and regulatory parameters of histone modifications. *J Cell Biochem* 30-31(suppl):203-213.
- Dyck JA, Maul GG, Miller WH, Chen JD, Kakizuka A, Evans RM. 1994. A novel macromolecular structure is a target of the promyelocyte-retinoic acid receptor oncoprotein. *Cell* 76:333-343.
- Giese K, Kingsley C, Kirshner JR, Grosschedl R. 1995. Assembly and function of a TCR alpha enhancer complex is dependent on LEF-1-induced DNA bending and multiple protein-protein interactions. *Genes Dev* 9:995-1008.
- Grande MA, van der Kraan I, van Steensel B, Schul W, de The H, van der Voort HT, de Jong L, van Driel R. 1996. PML-containing nuclear bodies: their spatial distribution in relation to other nuclear components. *J Cell Biochem* 63:280-291.

- Guo B, Odgren PR, van Wijnen AJ, Last TJ, Nickerson J, Penman S, Lian JB, Stein JL, Stein GS. 1995. The nuclear matrix protein NMP-1 is the transcription factor YY1. *Proc Natl Acad Sci USA* 92:10526–10530.
- Htun H, Barsony J, Renyi I, Gould DL, Hager GL. 1996. Visualization of glucocorticoid receptor translocation and intranuclear organization in living cells with a green fluorescent protein chimera. *Proc Natl Acad Sci USA* 93:4845–4850.
- Iborra FJ, Jackson DA, Cook PR. 1998. The path of transcripts from extra-nucleolar synthetic sites to nuclear pores: transcripts in transit are concentrated in discrete structures containing SR proteins. *J Cell Sci* 111 (pt 15):2269–2282.
- Imbalzano AN. 1998. Energy-dependent chromatin remodelers: complex complexes and their components. *Crit Rev Eukaryot Gene Expr* 8:225–255.
- Ito T, Bulger M, Pazin MJ, Kobayashi R, Kadonaga JT. 1997. ACF, an ISWI-containing and ATP-using chromatin assembly and remodeling factor. *Cell* 90:145–155.
- Jackson DA, Iborra FJ, Manders EM, Cook PR. 1998. Numbers and organization of RNA polymerases, nascent transcripts, and transcription units in HeLa nuclei [published erratum appears in *Mol Biol Cell* 1998; 9:2698]. *Mol Biol Cell* 9:1523–1536.
- Kimura H, Tao Y, Roeder RG, Cook PR. 1999. Quantitation of RNA polymerase II and its transcription factors in an HeLa cell: little soluble holoenzyme but significant amounts of polymerases attached to the nuclear substructure. *Mol Cell Biol* 19:5383–5392.
- Kwon H, Imbalzano AN, Khavari PA, Kingston RE, Green MR. 1994. Nucleosome disruption and enhancement of activator binding by a human SWI/SNF complex. *Nature* 370:477–481.
- Lamond AI, Earnshaw WC. 1998. Structure and function in the nucleus. *Science* 280:547–553.
- Leonhardt H, Rahn HP, Cardoso MC. 1998. Intranuclear targeting of DNA replication factors. *J Cell Biochem* 30–31(suppl):243–249.
- Ma H, Siegel AJ, Berezney R. 1999. Association of chromosome territories with the nuclear matrix. Disruption of human chromosome territories correlates with the release of a subset of nuclear matrix proteins. *J Cell Biol* 146:531–542.
- Mancini MG, Liu B, Sharp ZD, Mancini MA. 1999. Subnuclear partitioning and functional regulation of the Pit-1 transcription factor. *J Cell Biochem* 72:322–338.
- McNeil S, Guo B, Stein JL, Lian JB, Bushmeyer S, Seto E, Atchison ML, Penman S, van Wijnen AJ, Stein GS. 1998. Targeting of the YY1 transcription factor to the nucleolus and the nuclear matrix in situ: the C-terminus is a principal determinant for nuclear trafficking. *J Cell Biochem* 68:500–510.
- Merriman HL, van Wijnen AJ, Hiebert S, Bidwell JP, Fey E, Lian J, Stein J, Stein GS. 1995. The tissue-specific nuclear matrix protein, NMP-2, is a member of the AML/CBF/PEBP2/runt domain transcription factor family: interactions with the osteocalcin gene promoter. *Biochemistry* 34:13125–13132.
- Nickerson JA, Blencowe BJ, Penman S. 1995. The architectural organization of nuclear metabolism. In: Berezney R, Jeon KW, editors. *Structural and functional organization of the nuclear matrix*. San Diego: Academic Press. p 67–123.
- Ogawa E, Inuzuka M, Maruyama M, Satake M, Naito-Fujimoto M, Ito Y, Shigesada K. 1993. Molecular cloning and characterization of PEBP2 β , the heterodimeric partner of a novel *Drosophila* runt-related DNA binding protein PEBP2 α . *Virology* 194:314–331.
- Peterson CL, Zhao Y, Chait BT. 1998. Subunits of the yeast SWI/SNF complex are members of the actin-related protein (ARP) family. *J Biol Chem* 273:23641–23644.
- Pombo A, Cook PR. 1996. The localization of sites containing nascent RNA and splicing factors. *Exp Cell Res* 229:201–203.
- Rogaia D, Grignani F, Carbone R, Riganelli D, LoCoco F, Nakamura T, Croce CM, Di Fiore PP, Pelicci PG. 1997. The localization of the HRX/ALL1 protein to specific nuclear subdomains is altered by fusion with its eps15 translocation partner. *Cancer Res* 57:799–802.
- Silver PA, Keegan LP, Ptashine M. 1984. Amino terminus of the yeast GAL4 gene product is sufficient for nuclear localization. *Proc Natl Acad Sci USA* 81:5951–5955.
- Smith KP, Moen PT, Wydner KL, Coleman JR, Lawrence JB. 1999. Processing of endogenous pre-mRNAs in association with SC-35 domains is gene specific. *J Cell Biol* 144:617–629.
- Stenoien D, Sharp ZD, Smith CL, Mancini MA. 1998. Functional subnuclear partitioning of transcription factors. *J Cell Biochem* 70:213–221.
- Tang L, Guo B, Javed A, Choi J-Y, Hiebert S, Lian JB, van Wijnen AJ, Stein JL, Stein GS, Zhou GW. 1999. Crystal structure of the nuclear matrix targeting signal of the transcription factor AML-1/PEBP2 α B/CBFA2. *J Biol Chem* 274:33580–33586.
- Tang Y, Getzenberg RH, Vietmeier BN, Stallcup MR, Eggert M, Renkawitz R, DeFranco DB. 1998a. The DNA-binding and τ 2 transactivation domains of the rat glucocorticoid receptor constitute a nuclear matrix targeting signal. *Mol Endocrinol* 12:1420–1431.
- Tang L, Guo B, van Wijnen AJ, Lian JB, Stein JL, Stein GS, Zhou GW. 1998b. Preliminary crystallographic study of the glutathione S-transferase fused with the nuclear matrix targeting signal of the transcription factor AML-1/CBF α 2. *J Struct Biol* 123:83–85.
- Ullman KS, Powers MA, Forbes DJ. 1997. Nuclear export receptors: from importin to exportin. *Cell* 90:967–970.
- van Steensel B, Jenster G, Damm K, Brinkmann AO, van Driel R. 1995. Domains of the human androgen receptor and glucocorticoid receptor involved in binding to the nuclear matrix. *J Cell Biochem* 57:465–478.
- Wei X, Samarabandu J, Devdhar RS, Siegel AJ, Acharya R, Berezney R. 1998. Segregation of transcription and replication sites into higher order domains. *Science* 281:1502–1505.
- Wei X, Somanathan S, Samarabandu J, Berezney R. 1999. Three-dimensional visualization of transcription sites and their association with splicing factor-rich nuclear speckles. *J Cell Biol* 146:543–558.
- Workman JL, Kingston RE. 1998. Alteration of nucleosome structure as a mechanism of transcriptional regulation. *Annu Rev Biochem* 67:545–579.
- Yano T, Nakamura T, Blechman J, Sorio C, Dang CV, Geiger B, Canaani E. 1997. Nuclear punctate distribution of ALL-1 is conferred by distinct elements at the N terminus of the protein. *Proc Natl Acad Sci USA* 94:7286–7291.

- Zeng C, van Wijnen AJ, Stein JL, Meyers S, Sun W, Shopland L, Lawrence JB, Penman S, Lian JB, Stein GS, Hiebert SW. 1997. Identification of a nuclear matrix targeting signal in the leukemia and bone-related AML/CBF α transcription factors. *Proc Natl Acad Sci USA* 94:6746–6751.
- Zeng C, McNeil S, Pockwinse S, Nickerson JA, Shopland L, Lawrence JB, Penman S, Hiebert SW, Lian JB, van Wijnen AJ, Stein JL, Stein GS. 1998. Intranuclear targeting of AML/CBF α regulatory factors to nuclear matrix-associated transcriptional domains. *Proc Natl Acad Sci USA* 95:1585–1589.
- Zhang DE, Hetherington CJ, Meyers S, Rhoades KL, Larson CJ, Chen HM, Hiebert SW, Tenen DG. 1996. CCAAT enhancer-binding protein (C/EBP) and AML1 (CBF α 2) synergistically activate the macrophage colony-stimulating factor receptor promoter. *Mol Cell Biol* 16:1231–1240.
- Zlatanova JS and van Holde KE. 1992. Chromatin loops and transcriptional regulation. *Crit Rev Eukaryot Gene Expr* 2:211–224.